Hydrogen Peroxide Oxygenation of Furan-2-carbaldehyde via an Easy, Green Method

Viktor Zvarych,^{[†](#page-2-0)}® Anna Nakonechna,[†]® Mykhailo Marchenko,[‡](#page-2-0)® Oleksii Khudyi,[‡]® Vira Lubenets,[†] Lidiia Khuda,^{[‡](#page-2-0)}® Olga Kushniryk,^{‡®} and Volodymyr Novikov^{[*](#page-2-0),†}

 † Department of Technology of Biologically Active Substances, Pharmacy and Biotechnology, Lviv Polytechnic National University, Stepan Bandera Street 12, Lviv 79013, Ukraine

 ‡ Department of Biochemistry and Biotechnology, Yuriy Fedkovych Chernivtsi National University, Kotsyubynsky Street 2, Chernivtsi 58012, Ukraine

ABSTRACT: Derivatives of 2(5H)-furanone (γ-crotonolactone) are important intermediate synthetic products with a wide range of biological effects that have become widely used in the pharmaceutical industry, medicine, and veterinary medicine, in particular in the prevention and treatment of fish diseases. However, the environmental issue of obtaining these compounds while reducing the negative impact on the surrounding environment remains relevant. This article describes for the first time a method of γ-crotonolactone synthesis that is based on the concept of green chemistry. Synthesis is carried out under mild conditions using nontoxic reagents by furfural oxidation. For the first time, a mixture of hydrogen peroxide and acetic acid was used for the oxidation of furfural in a ratio of 1:0.05. A mixture of organic acids (succinic, maleic, fumaric, formic, and cinnamic acids), obtained as a byproduct in the synthesis of γ-crotonolactone, can be used as a highly effective, ecofriendly organic fertilizer or in a preparation with a stimulating effect.

KEYWORDS: furan-2-carbaldehyde, 2(5H)-furanone (γ-crotonolactone), green chemistry, hydrogen peroxide, acetic acid

ENTRODUCTION

The use of compounds of the furan series, namely, derivatives of 2(5H)-furanone (γ-crotonolactone or 2-butenolide), in synthetic organic chemistry and in the national economy have been known for a long time.^{[1](#page-2-0),[2](#page-2-0)} Compounds of this class are found in beer, kvass, fermented foods, and especially in contaminated feeds exposed to mold.^{[3,4](#page-2-0)} They attract considerable attention as growth regulators of plants; 5 substances that inhibit tumor growth in organisms, as they are inhibitors of cell metamorphosis;^{[6](#page-2-0)} and promoters of the growth of industrial fish cultures.^{[7](#page-2-0)} 2(5H)-Furanones are known as compounds with antifungal and antibacterial activities.^{[8](#page-2-0)−[10](#page-2-0)}

Under the influence of γ -crotonolactone, the contents of thiol groups in the intestinal mucosa increase because of increased biosynthetic processes in the gut associated with increased secretion of specific enzymes. The experiments for the determination of the total proteolytic activity of fish intestine exposed to γ -crotonolactone demonstrate this very clearly. The total proteolytic activity, characterizing the number of enzymes in the intestinal mucosa, increases significantly (by 24%) in the second day and continues to remain at a high level for 3 days. On the 30th day, it grows up to 30%. It is noted that the proteolytic activity of the intestine continues to remain high even after the end of γ crotonolactone addition to the feed. $2(5H)$ -Furanone does not have a negative effect on the fish body; on the contrary, this drug helps to suppress the growth of pathogenic microflora in the intestine. $11,12$ $11,12$

At the same time, formic, fumaric, maleic, succinic, and cinnamic acids as a product of the furfural oxidation are used in almost all fields of the industrial and food chemistry. Formic acid and its salts are used in the feed industry, textile dyeing and finishing, food additives, grass silage, natural rubber, leather tanning, etc.¹³ Fumaric acid is nontoxic and versatility reagent, that used in the resin industry, green chemistry as an eco-friendly catalyst, and in the food industry as a nutritional additive and acidulant.^{[14](#page-2-0)} Maleic acid is not so widespread like fumaric acid. Maleic acid is used for the production of maleinate resins, copolymers, and generally as a chemical precursor of fumaric acid and maleic anhydride.^{[15](#page-2-0)} Moreover, cinnamic acid and its derivatives are known as natural occurring products.^{[16](#page-2-0)} They were found in all green plants.^{[17](#page-2-0)} Cinnamic acids are also used in medicinal chemistry as a perspective objects with a different useful spectrum of activities.^{[18](#page-2-0)}

Another promising direction for the use of $2(5H)$ -furanone is fish breeding, which is particularly widespread in Central and Eastern Europe as well as in Asia. Drugs based on $2(5H)$ furanone inhibit the development of cyanobacteria, which prevents fish death in ponds. In particular, there is a positive result from the use of drugs based on γ-crotonolactone and from the use of a mixture of the above-mentioned organic acids in the cultivation of fish in natural reservoirs and in recirculating aquaculture systems, which stimulate the development of a natural fodder base in fish-breeding ponds.¹⁹ This allows the use of such drugs in the intensive cultivation of live feeds, in particular those of freshwater planktonic crustaceans such as daphniids. These organisms are widely used both in

Scheme 1. Oxidation Mechanism of Furan-2-carbaldehyde

ornamental aquaculture and in the larviculture of many commercial fish species. 20

Therefore, the aim of this study was to improve the synthesis of 2(5H)-furanone using nontoxic and environmentally friendly hydrogen peroxide oxygenation in the presence of a catalytic amount of acetic acid in water.

■ MATERIALS AND METHODS

Chemicals. All analytical-grade chemicals were obtained from Sigma-Aldrich and were used without any further purification. Double-distilled water was used in this study.

General Experimental Details. All melting points were determined in open capillary tubes on a Boetius apparatus and are uncorrected. The $^1\mathrm{H}$ NMR and $^{13}\mathrm{C}$ NMR spectra were recorded on a Bruker Avance DRX-500 spectrometer (500 MHz) in DMSO- d_{6} ; the chemical shifts were measured relative to tetramethylsilane. Mass spectra were recorded on an Agilent 1100 Series G1956B LC/MSD SL LCMS system using electrospray ionization at atmospheric pressure (70 eV). UV spectra were recorded on a SPECORD M 40 spectrophotometer. IR spectra were recorded on a SPECORD M 80 spectrophotometer in tablets with KBr. Elemental analysis was carried out on a PerkinElmer 2400 CHN analyzer. Separate determination was carried out using an HPP4001 liquid chromatograph with two serially coupled glass columns (150 \times 3.3 mm) packed with the Separon SC-X C-18 adsorbent (0.6% acetic acid as the mobile phase, flow rate of 0.3 cm³/min). Monitoring of the reactions was performed by TLC on Silufol UV 254 plates.

Oxidation of Furfural. The oxidation reaction with hydrogen peroxide was carried out in a three-necked round-bottom glass flask equipped with a reflux condenser and a mechanical stirrer. The flask was loaded under stirring with the furfural (13.0 mL, 0.160 mol) and water (50 mL), and a mixture of 37.5% H_2O_2 (14.7 mL, 0.192 mol) with acetic acid (0.55 mL, 0.0096 mol) was slowly added. The reaction mixture was kept under constant stirring for 24 h at a temperature below 60 °C.

After completion of the reaction, water was removed under reduced pressure. The light-yellow precipitate of organic acids was filtered off. 2(5H)-Furanone was obtained from the filtrate by vacuum distillation at 12 mmHg. The yield of 2(5H)-furanone was 71%.

2(5H)-Furanone (1). Colorless to slightly light-yellow liquid, bp 85−88 °C at 12 mmHg. ¹H NMR (500 MHz, CDCl₃) *δ* 4.88 (dd, ⁴J $= 2.2 \text{ Hz}, \frac{3}{J} = 1.7 \text{ Hz}, 2\text{H}, 5\text{-H}, 6.10 \text{ (dt, } 4J = 2.2 \text{ Hz}, \frac{3}{J} = 5.8 \text{ Hz}, 1\text{H},$ 3-H), 7.58 (dt, 4 J = 1.7 Hz, 3 J = 5.8 Hz, 1H, 4-H). ¹³C NMR (125 MHz, CDCl₃,) δ 71.9, 121.1, 152.2, 173.7. IR (KBr, cm⁻¹): 1739, 1781 (C=O), 1053-1205 (-C-O-C−). Anal. calcd for C₄H₄O₂: C, 57.14; H, 4.80. Found: C, 57.10; H, 4.71. MS (EI) m/z 85.11 [M + H]+ .

The calculated and found data for succinic, maleic, fumaric, formic, and cinnamic acids were confirmed by the literary data.

■ RESULTS AND DISCUSSION

Catalytic oxidation of furan-2-carbaldehyde (furfural) attracts the attention of many researchers. This aldehyde is derived from vegetable wastes $^{21-23}$ $^{21-23}$ $^{21-23}$ $^{21-23}$ $^{21-23}$ and is available on the market as a chemical reagent. The furfural molecule contains several reaction centers, which implies a wide variety of syntheses. Projects on the oxidation of furfural by molecular oxygen after photoinitiation²⁴ and in conditions of heterogeneous catalysis, including through the use of compounds of transitional and noble metals $(Cu(OAc))_2$, $Mn(OAc))_2$, $Pd(OAc))_2$, AgOAc,

FeSO₄, RuCl₃, NiCl₂, V₂O₅, Co(NO₃)₂, TiO₂–ZrO₂, and $Pb(OAc)_2$) are well-known.^{[25](#page-2-0)−[34](#page-3-0)} The oxidation of furfural with hydrogen peroxide in the presence of a photogenic iron catalyst in the form of a complex $[FeCp(C₆H₅R)]PF₆$ (R = H, Cl, or $CH₃$) was also reported.^{[35](#page-3-0)} However, insufficient attention was paid to the homogeneous reactions of furfural with aqueous hydrogen peroxide in the works mentioned above.

At the same time, it is known that in the process of furfural oxidation with hydrogen peroxide (Scheme 1), depending on the reaction conditions and the catalyst type, various reactive compounds are formed, $36,37$ $36,37$ which provide the means for the implementation of various synthetic pathways on the basis of the reactions of furfural with aqueous H_2O_2 and catalysts.

All of the methods given above are multistage and include the use of hard-to-reach and in some cases toxic materials, such as organic solvents (dichloroethane and chloroform) and complex salts of heavy and transition metals, such as Cu^{2+} , Mn^{2+} , Pd²⁺, Ag⁺, Ru³⁺, Ni²⁺, V⁵⁺, and Co²⁺, in the role of catalyst. These catalysts are expensive and have a negative impact on the environment.

Therefore, in this work, a method of γ-crotonolactone synthesis was designed on the basis of the concept of green chemistry; namely, the oxidation of furfural was carried out in an aqueous solution without the use of organic solvents, and for the first time, an aqueous solution of hydrogen peroxide was used in the role of an oxidizer with a small amount of acetic acid as a promoter of the reaction. Oxidation was carried out for 24 h with vigorous stirring and at a temperature below 60 °C. The process was controlled by UV spectroscopy (residual aldehyde) and TLC (acid formation). The compositions of the reaction products were determined by high-performance liquid chromatography (HPLC) using authentic samples. As a result, the target product, γ crotonolactone (1), was obtained with a yield of ∼71% and a light-yellow crystalline precipitate (Scheme 2).

Using HPLC, it was determined that the crystalline precipitate contained mainly a mixture of organic acids,

Scheme 2. Oxidation of Furan-2-carbaldehyde with H_2O_2 and Acetic Acid in Water

namely, succinic (2) , maleic (3) , fumaric (4) , formic (5) , and cinnamic acids (6). This mixture of organic acids can be used in agriculture as nontoxic, ecologically pure organic fertilizer.

Thus, using the concept of green chemistry, γ-crotonolactone, an important intermediate synthetic product with a wide range of biological effects, was synthesized by the oxidation of furfural under soft conditions and using nontoxic reagents. For the oxidation of furfural, a mixture of hydrogen peroxide and acetic acid was used in a ratio of 1:0.05 for the first time. The byproduct was a mixture of organic acids that can be used in the national economy as a highly effective, environmentally friendly organic fertilizer.

■ AUTHOR INFORMATION

Corresponding Author

*Tel.: +380-67-6743067. E-mail: [volodymyr.p.novikov@lpnu.](mailto:volodymyr.p.novikov@lpnu.ua) [ua](mailto:volodymyr.p.novikov@lpnu.ua).

ORCID[®]

Viktor Zvarych: [0000-0003-3036-0050](http://orcid.org/0000-0003-3036-0050) Anna Nakonechna: [0000-0003-3539-0160](http://orcid.org/0000-0003-3539-0160) Mykhailo Marchenko: [0000-0001-6104-0119](http://orcid.org/0000-0001-6104-0119) Oleksii Khudyi: [0000-0001-5652-0900](http://orcid.org/0000-0001-5652-0900) Vira Lubenets: [0000-0001-6189-0084](http://orcid.org/0000-0001-6189-0084) Lidiia Khuda: [0000-0002-1098-7537](http://orcid.org/0000-0002-1098-7537)

Olga Kushniryk: [0000-0003-1001-6711](http://orcid.org/0000-0003-1001-6711)

Volodymyr Novikov: [0000-0002-0485-8720](http://orcid.org/0000-0002-0485-8720)

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

The authors would like to express their gratitude to the Ministry of Education and Science of Ukraine (Scientific Research Project Number 0116U004138) for financial support.

■ REFERENCES

(1) Rossi, R.; Lessi, M.; Manzini, C.; Marianetti, G.; Bellina, F. Synthesis and biological properties of 2(5H)-furanones featuring bromine atoms on the heterocyclic ring and/or brominated substituents. Curr. Org. Chem. 2017, 21, 964−1018.

(2) Rao, Y. S. Recent advances in the chemistry of unsaturated lactones. Chem. Rev. 1976, 76, 625−694.

(3) Slaughter, J. C. The naturally occurring furanones: Formation and function from pheromone to food. Biol. Rev. Camb. Philos. Soc. 1999, 74, 259−276.

(4) Scholtes, C.; Nizet, S.; Collin, S. How Sotolon Can Impart a Madeira Off-Flavor to Aged Beers. J. Agric. Food Chem. 2015, 63, 2886−2892.

(5) Light, M. E.; Burger, B. V.; Staerk, D.; Kohout, L.; Van Staden, J. Butenolides from Plant-Derived Smoke: Natural Plant-Growth Regulators with Antagonistic Actions on Seed Germination. J. Nat. Prod. 2010, 73, 267−269.

(6) Dobretsov, S.; Teplitski, M.; Paul, V. Mini-review: Quorum sensing in the marine environment and its relationship to biofouling. Biofouling 2009, 25, 413−427.

(7) Nemchenko, V. V.; Ivanova, N. P. Growth regulators for dressing of corn seed. Khim. Sel'sk. Khoz. 1991, 1, 91−93; Chem. Abstr. 1991, 115, 44136.

(8) Sakurai, K.; Matsumoto, H.; Adachi, J. Antifungal studies on drugs. I. Antifungal activity of five-membered lactone derivatives. Yakugaku Zasshi 1968, 88, 919−924.

(9) Paulitz, T.; Nowak-Thompson, B.; Gamard, P.; Tsang, E.; Loper, J. A novel antifungal furanone from Pseudomonas aureofaciens, a biocontrol agent of fungal plant pathogens. J. Chem. Ecol. 2000, 26, 1515−1524.

(10) Šenel, P.; Tichotová, L.; Votruba, I.; Buchta, V.; Špulák, M.; Kuneš, J.; Nobilis, M.; Krenk, O.; Pour, M. Antifungal 3,5disubstituted furanones: From 5-acyloxymethyl to 5-alkylidene derivatives. Bioorg. Med. Chem. 2010, 18, 1988−2000.

(11) Markad, A.; Rane, M. Use of Probiotics in Aquaculture. Int. J. Sci. Technoledge 2015, 3, 1−6.

(12) Natrah, F. M. I.; Defoirdt, T.; Sorgeloos, P.; Bossier, P. Disruption of Bacterial Cell-to-Cell Communication by Marine Organisms and its Relevance to Aquaculture. Mar. Biotechnol. 2011, 13, 109−126.

(13) Hietala, J.; Vuori, A.; Johnsson, P.; Pollari, I.; Reutemann, W.; Kieczka, H. Formic Acid. In Ullmann's Encyclopedia of Industrial Chemistry; Wiley: Weinheim, Germany, 2016; [DOI: 10.1002/](http://dx.doi.org/10.1002/14356007.a12_013.pub3) [14356007.a12_013.pub3.](http://dx.doi.org/10.1002/14356007.a12_013.pub3)

(14) Das, R. K.; Brar, S. K.; Verma, M. Fumaric Acid: Production and Application Aspects. In Platform Chemical Biorefinery: Future Green Chemistry, 1st ed.; Brar, S. K., Sarma, S. J., Pakshirajan, K., Eds.; Elsevier, 2016; pp 133−157.

(15) Lohbeck, K.; Haferkorn, H.; Fuhrmann, W.; Fedtke, N. Maleic and Fumaric Acids. In Ullmann's Encyclopedia of Industrial Chemistry, 7th ed.; Bellussi, G., Bohnet, M., Bus, J., Drauz, K., Greim, H., Jäckel, K.-P., Karst, U., Kleemann, A., Kreysa, G., Laird, T., Meier, W., Ottow, E., Röper, M., Scholtz, J., Sundmacher, K., Ulber, R., Wietelmann, U., Eds.; Wiley: Weinheim, Germany, 2000; Vol. 40.

(16) Clifford, M. N. Chlorogenic acids and other cinnamates − Nature, occurrence and dietary burden. J. Sci. Food Agric. 1999, 79, 362−372.

(17) Lichtenthaler, H. K.; Schweiger, J. Cell wall bound ferulic acid, the major substance of the blue-green fluorescence emission of plants. J. Plant Physiol. 1998, 152, 272−282.

(18) Guzman, J. D. Natural cinnamic acids, synthetic derivatives and hybrids with antimicrobial activity. Molecules 2014, 19, 19292−19349. (19) Tsen, L. N.; Suleimanyan, V. S. Effect of crotonolactone on the

roe and fry of the rainbow trout. Rybn. Khoz. 1982, 5, 37−42; Chem. Abstr. 1982, 97, 22625.

(20) Dhert, P.; Lim, L. C.; Candreva, P.; Van Duffel, H.; Sorgeloos, P. Possible applications of modern fish larviculture technology to ornamental fish production. Aquarium Sci. Conserv. 1997, 1, 119−128.

(21) Zeitsch, K. J. Furfural Processes. In The chemistry and technology of furfural and its many by-products, 1st ed.; Zeitsch, K. J., Ed.; Elsevier Science and Technology, 2000; Vol. 13, pp 36−74.

(22) Kamm, B.; Kamm, M.; Schmidt, M.; Hirth, T.; Schulze, M. Lignocellulose based Chemical Products and Product Family Trees. In Biorefineries−Industrial Processes and Products: Status Quo and Future Directions, 1st ed.; Kamm, B., Gruber, P. R., Kamm, M., Eds.; Wiley: Weinheim, Germany, 2006; Vol. 2, pp 97−149.

(23) Chheda, J. N.; Huber, G. W.; Dumesic, J. A. Liquid Phase Catalytic Processing of Biomass Derived Oxygenated Hydrocarbons to Fuels and Chemicals. Angew. Chem., Int. Ed. 2007, 46, 7164−7183.

(24) Borghei, S. M.; Hosseini, S. N. Comparison of furfural degradation by different photooxidation methods. Chem. Eng. J. 2008, 139, 482−488.

(25) Shi, S.; Guo, H.; Yin, G. Synthesis of maleic acid from renewable resources: Catalytic oxidation of furfural in liquid media with dioxygen. Catal. Commun. 2011, 12, 731−733.

(26) Pinna, F.; Olivo, A.; Trevisan, V.; Menegazzo, F.; Signoretto, M.; Manzoli, M.; Boccuzzi, F. The effects of gold nanosize for the exploitation of furfural by selective oxidation. Catal. Today 2013, 203, 196−201.

(27) Li, F.; Lu, T.; Chen, B.; Huang, Z.; Yuan, G. Pt nanoparticles over TiO₂−ZrO₂ mixed oxide as multifunctional catalysts for an integrated conversion of furfural to 1,4-butanediol. Appl. Catal., A 2014, 478, 252−258.

(28) Verdeguer, P.; Merat, N.; Gaset, A. Lead/platinum on charcoal as catalyst for oxidation of furfural. Effect of main parameters. Appl. Catal., A 1994, 112, 1−11.

(29) Menegazzo, F.; Signoretto, M.; Pinna, F.; Manzoli, M.; Aina, V.; Cerrato, G.; Boccuzzi, F. Oxidative esterification of renewable furfural

Journal of Agricultural and Food Chemistry Article and The State Art

(30) Signoretto, M.; Menegazzo, F.; Contessotto, L.; Pinna, F.; Manzoli, M.; Boccuzzi, F. Au/ZrO₂ an efficient and reusable catalyst for the oxidative esterification of renewable furfural. Appl. Catal., B 2013, 129, 287−293.

(31) Thorton, M. M.; Malte, P. C.; Crittenden, A. L. Oxidation of furan and furfural in a well-stirred reactor. Symp. Combust., [Proc.] 1988, 21, 979−989.

(32) Murthy, M. S.; Rajamani, K. Kinetics of vapour phase oxidation of furfural on vanadium catalyst. Chem. Eng. Sci. 1974, 29, 601−609.

(33) Badovskaya, L. A.; Latashko, V. M.; Poskonin, V. V.; Grunskaya, E. P.; Tyukhteneva, Z. I.; Rudakova, S. G.; Pestunova, S. A.; Sarkisyan, A. V. Catalytic Oxidation of Furan and Hydrofuran Compounds. 7. Production of 2(5H)-Furanone by Oxidation of Furfural with Hydrogen Peroxide and Some of Its Transformations in Aqueous Solutions. Chem. Heterocycl. Compd. 2002, 38, 1040−1048.

(34) Zhao, X.; Wang, L. Atmospheric Oxidation Mechanism of Furfural Initiated by Hydroxyl Radicals. J. Phys. Chem. A 2017, 121, 3247−3253.

(35) Moulines, F.; Ruiz, J.; Astruc, D. Oxidation of furfural with $H₂O₂$ in the presence of a photogenerated iron catalyst. J. Organomet. Chem. 1988, 340, C13−C14.

(36) Badovskaya, L. A.; Poskonin, V. V. Metal nature effect on catalytic reactions in furfural- H_2O_2 - H_2O -group V or VI d-metal salt systems in acid media. Kinet. Catal. 2015, 56, 164−172.

(37) Cao, R.; Liu, C.; Liu, L. A convenient synthesis of 2(5H) furanone. Org. Prep. Proced. Int. 1996, 28, 215−216.